

Genetics Primer – Genetics for Central Cancer Registries

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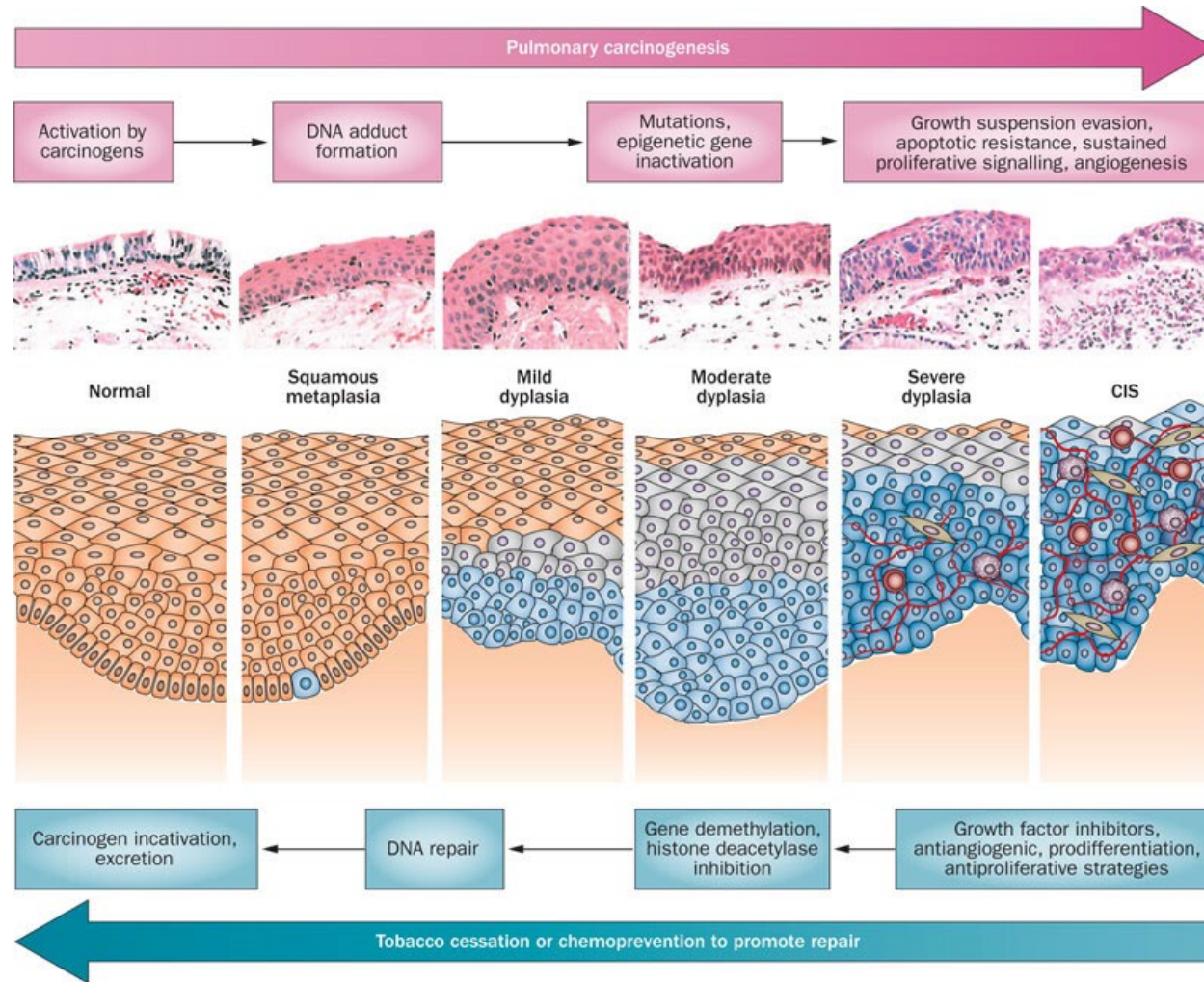
Topics to be Covered

- Why collect genomic tumor data?
- Genomic data routinely generated for clinical oncology
- A central registry approach to surveillance of genomic data
- Central registry infrastructure needed

Why Collect Genomic Tumor Data?

A Public Health Cancer Surveillance Imperative

Carcinogenesis: A Genomic Process



Keith, R., Miller, Y. Lung cancer chemoprevention: current status and future prospects. *Nat Rev Clin Oncol* 10, 334–343 (2013). <https://doi.org/10.1038/nrclinonc.2013.64>

NSCLC Treatment Before Genomics

2-drug platinum-based regimens

Stratification

Performance status
0-1 vs. 2

Weight loss in previous 6 months
<5% vs. ≥5%

Disease stage IIIB or IV

Presence or absence of brain metastases

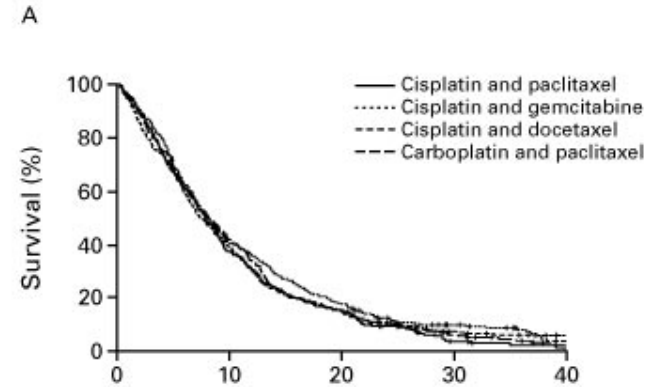
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→ **Arm A: Cisplatin + Paclitaxel**
Paclitaxel: 135 mg/m² over 24 hours, day 1
Cisplatin: 75 mg/m² day 2
3-week cycle

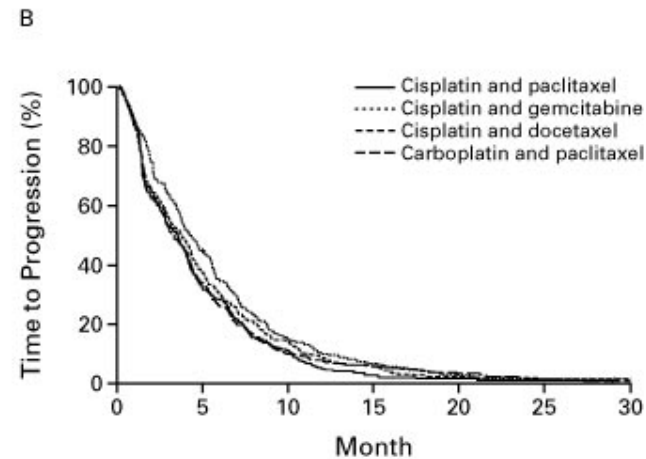
→ **Arm B: Cisplatin + Gemcitabine**
Gemcitabine: 1,000 mg/m² days 1,8,15
Cisplatin: 100 mg/m² day 1
4-week cycle

→ **Arm C: Cisplatin + Docetaxel**
Docetaxel: 75 mg/m² day 1
Cisplatin: 75 mg/m² day 1
3-week cycle

→ **Arm D: Carboplatin + Paclitaxel**
Paclitaxel: 225 mg/m² over 3 hours, day 1
Carboplatin: AUC 6.0 day 1
3-week cycle



Median survival
8 mo.

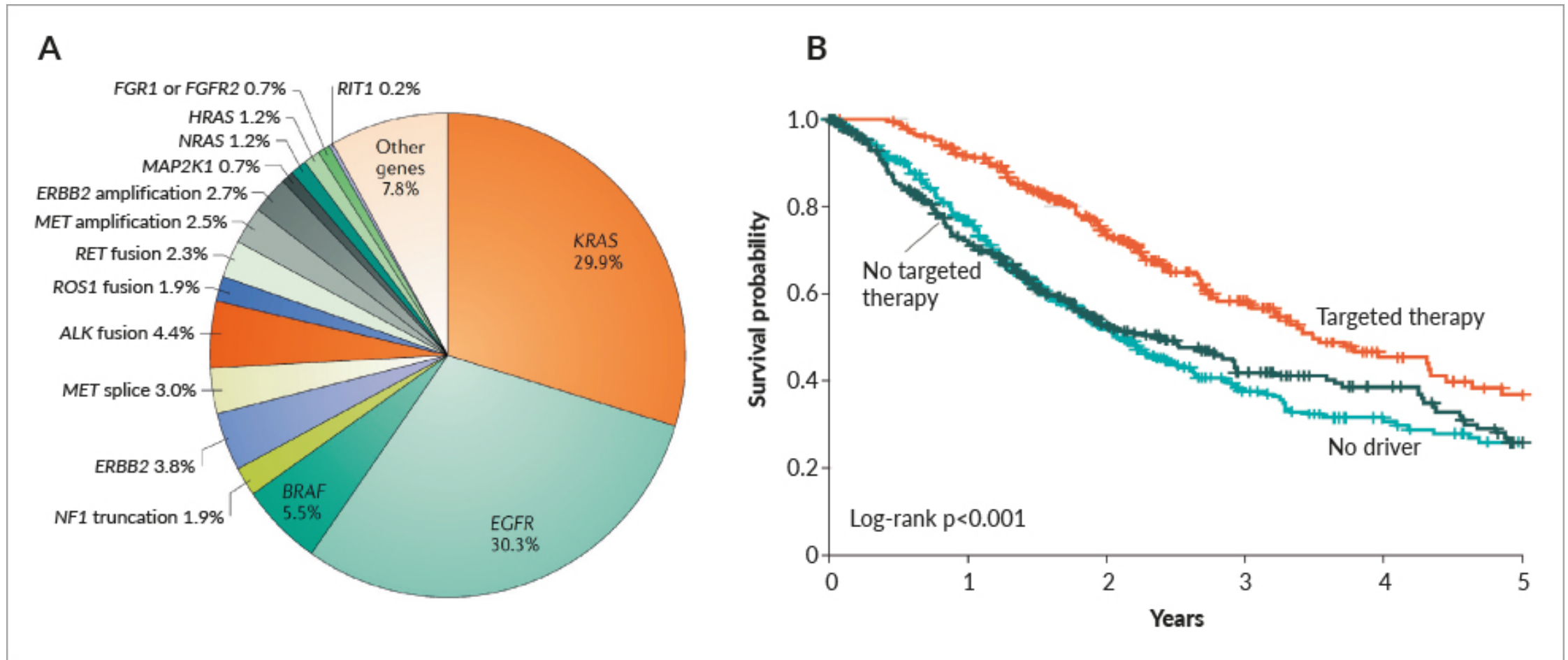


Response rate
19%

Median time to tumor progression
(TTP) 3.7 mo.

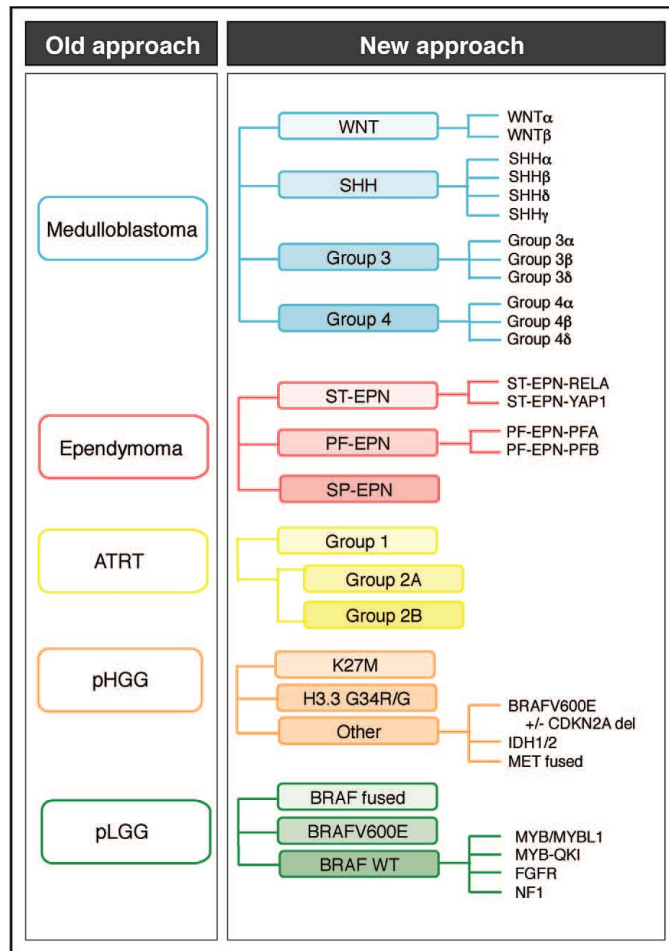
Schiller JH. NEJM 2002; 346:92-98

Current and Future Targets in NSCLC



Meisel 2020, Healthbook

Shifting Paradigm in Treatment of Pediatric Brain Tumors



- Medulloblastoma – Subgroup of Embryonal Tumors
- Ependymoma – A type of CNS Tumor
- Atypical Teratoid/Rhabdoid Tumors (ATRT) – Rhabdoid tumors of the CNS, common in very young children
- Pediatric High-Grade Glioma (pHGG) – heterogenous malignant tumors
- Pediatric Low-Grade Glioma (pLGG) – histologically diverse benign tumors of glial origin

Genomic Data Capture: A Public Health Imperative

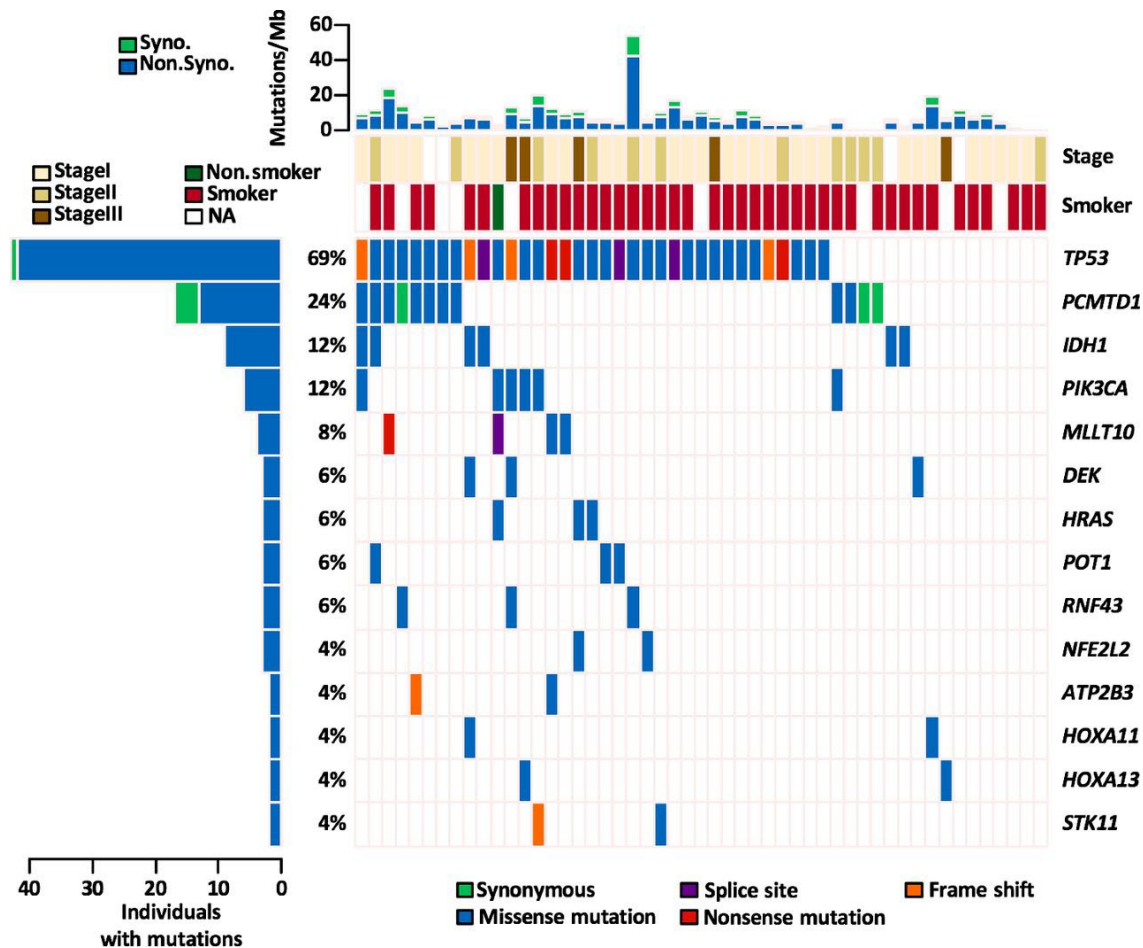
How do genomic variants impact treatment, treatment response, and survival in the population?

Do disparities exist in patients who have access to molecular testing and targeted therapy?

Do molecular profiles vary by geography, race/ethnicity, or socio-economic status?

Can genetic testing be used to identify cancer risk, diagnose cancer sooner or prevent cancer?

Appalachian KY vs TCGA Mutations in Whole Exome Sequencing of Squamous Cell Lung Cancer (N=51)



This study identified an increased percentage of IDH1 and PCMTD1 mutations in SQCC arising in the Appalachian KY residents versus TCGA, with population-specific implications for the personalized treatment of this disease.

EGFR Testing and Erlotinib Use in Non-Small Cell Lung Cancer Patients in Kentucky

Modeling Had EGFR Testing		
Variable	OR (95% CI)	P-Value
Age (ref = 75+)		0.0001
20–49	4.15 (2.17–7.91)	
50–64	1.76 (1.16–2.67)	
65–74	1.39 (0.98–1.98)	
Sex (ref = Male)		0.0142
Female	1.44 (1.08–1.93)	
Appalachian Status (ref = Non-Appalachia/Metro)		0.0011
Appalachian/Metro	0.67 (0.28–1.59)	
Appalachian/Non-Metro	0.51 (0.36–0.73)	
Non-Appalachian/Non-Metro	0.60 (0.40–0.89)	
Year of Diagnosis (ref = 2007)		<0.0001
2008	3.81 (0.43–34.68)	
2009	22.30 (3.00–165.41)	
2010	58.56 (8.12–422.26)	
2011	113.47 (15.81–814.21)	
Insurance (ref = Private)		<0.0001
Medicaid	0.19 (0.09–0.40)	
Medicare	0.61 (0.44–0.84)	
Smoking (ref = No)		0.0266
Yes	0.54 (0.32–0.91)	
Unknown	0.83 (0.42–1.66)	

OR = odds ratio; CI = confidence interval; (ref) = reference variable

<https://doi.org/10.1371/journal.pone.0237790.t003>

Modeling Receive Erlotinib		
Variable	OR (95% CI)	P-Value
Age (ref = 75+)		0.0077
20–49	2.05 (1.02–4.14)	
50–64	1.97 (1.31–2.95)	
65–74	1.56 (1.10–2.21)	
Sex (ref = Male)		0.0045
Female	1.49 (1.13–1.97)	
Insurance (ref = Private)		0.0074
Medicaid	0.55 (0.33–0.93)	
Medicare	0.63 (0.46–0.87)	
Poverty (ref = Low)		0.0081
Moderate	1.90 (1.24–2.91)	
High	1.84 (1.22–2.79)	
Very High	1.33 (0.85–2.09)	

OR = odds ratio; CI = confidence interval; (ref) = reference variable

<https://doi.org/10.1371/journal.pone.0237790.t004>

Larson KL, Huang B, Chen Q, Tucker T, Schuh M, et al. (2020) EGFR testing and erlotinib use in non-small cell lung cancer patients in Kentucky. PLOS ONE 15(8): e0237790. <https://doi.org/10.1371/journal.pone.0237790>

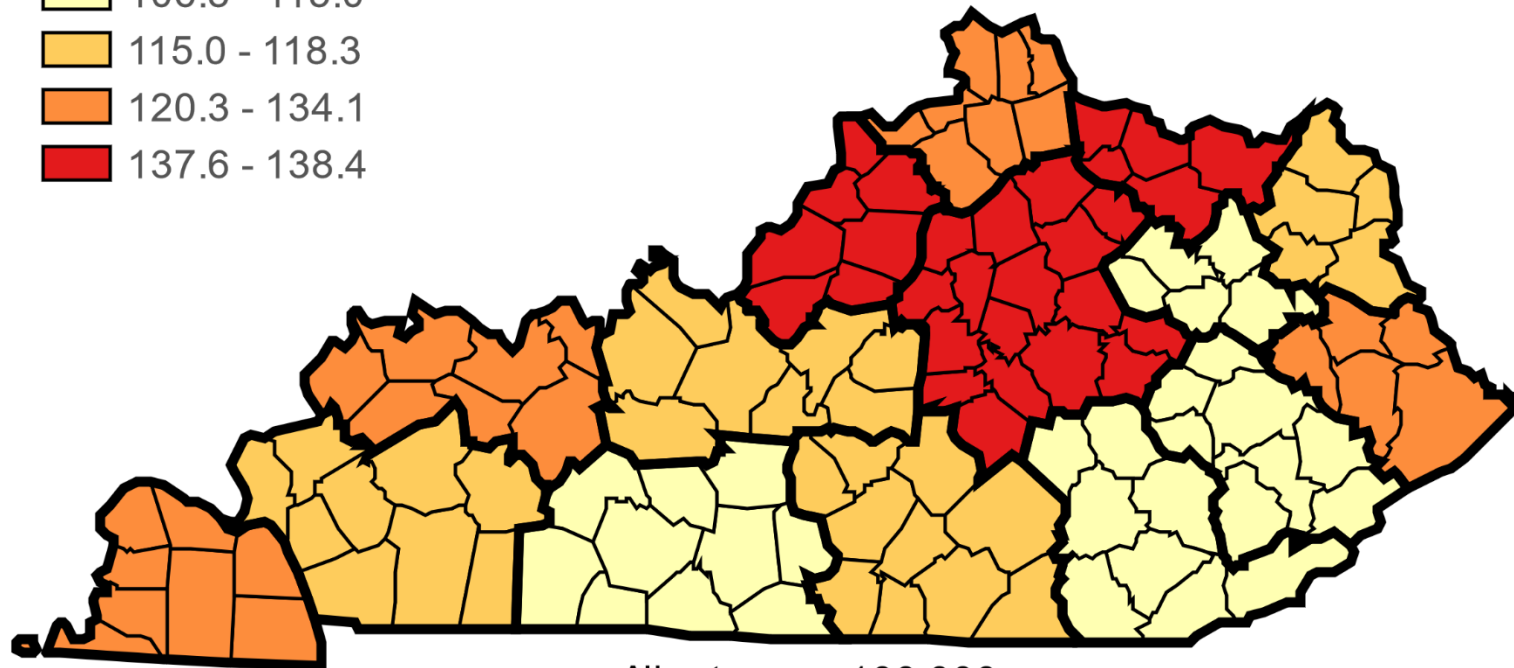
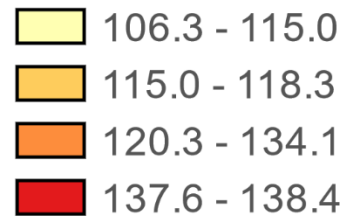
Age-Adjusted Invasive Cancer Incidence Rates in Kentucky

Breast, Female, 2014 - 2018

By Area Development District

Age-Adjusted to the 2000 U.S. Standard Million Population

Kentucky Rate: 128.0 / per 100,000



All rates per 100,000.

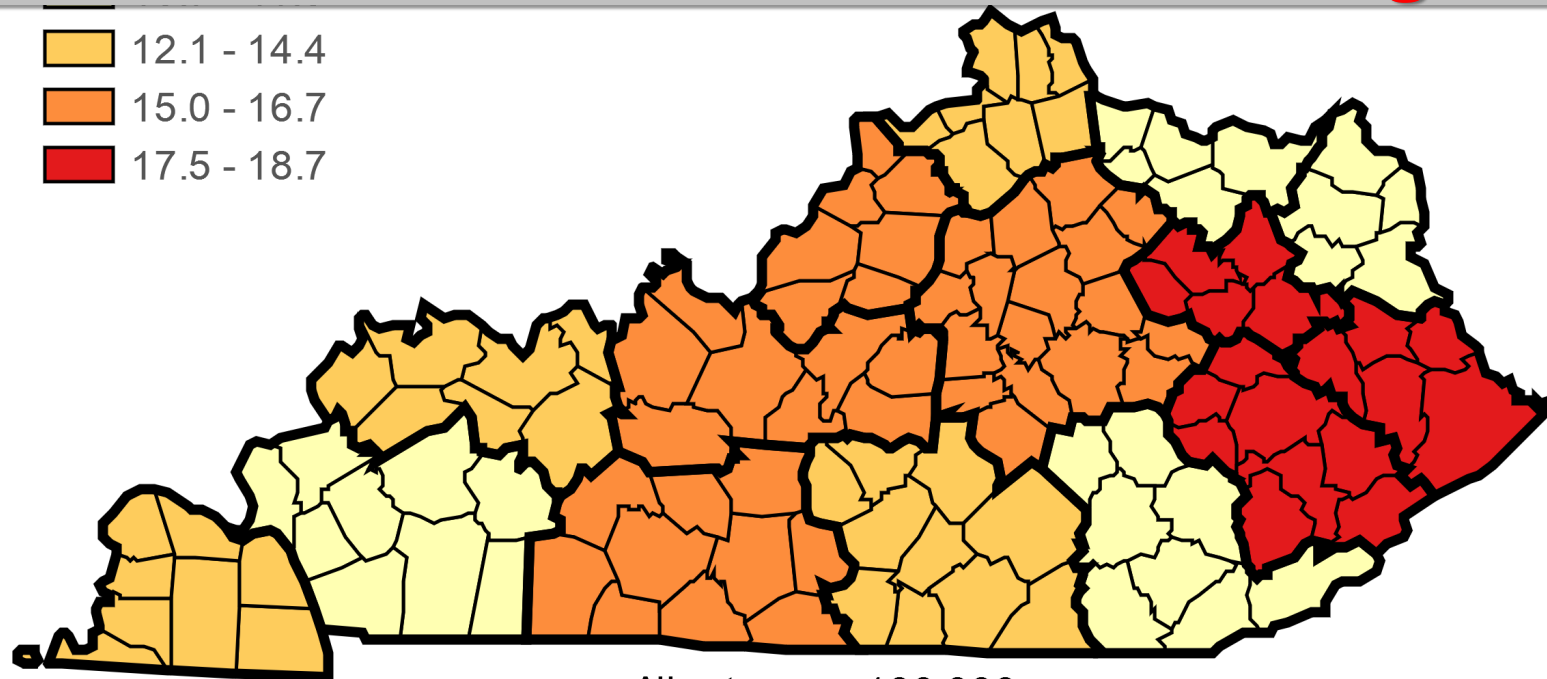
Data accessed July 26, 2022. Based on data released Nov 2021.

© 2022 Kentucky Cancer Registry.

Age-Adjusted Invasive Cancer Incidence Rates in Kentucky
Triple Negative (HR-/HER2-) - Breast, Female, 2014 - 2018
By Area Development District
Age-Adjusted to the 2000 U.S. Standard Million Population

Kentucky Rate: 14.7 / per 100,000

Do geographic variations also exist in incidence of cancer genomic subtypes?



All rates per 100,000.
Data accessed September 16, 2021. Based on data released Nov 2020.
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Genomic Data Generated for Clinical Oncology

New Challenges in Cancer Surveillance

Next Generation Sequencing (NGS) Multi-Gene Targeted Panels

Kentucky Cancer Registry (KCR) Cancer Genomics Data Sources

- Clinical NGS reports
- Research NGS Reports
Oncology Research
Information Exchange
Network (ORIEN)
- Pediatric Brain Tumor Study

Common Clinical NGS Service Providers in U.S.

- Guardant Health
- Foundation Medicine
- Caris Life Sciences
- Tempus
- Others

Current Gene List²

Genes with full coding exonic regions included in FoundationOne[®]CDx for the detection of substitutions, insertion-deletions (indels), and copy-number alterations (CNAs).

ABL1	ACVR1B	AKT1	AKT2	AKT3	ALK	ALOX12B	AMER1 (FAM123B)	APC
AR	ARAF	ARFRP1	ARID1A	ASXL1	ATM	ATR	ATRX	AURKA
AURKB	AXINI	AXL	BAP1	BARD1	BCL2	BCL2L1	BCL2L2	BCL6
BCOR	BCORL1	BRAF	BRCA1	BRCA2	BRD4	BRIP1	BTG1	BTG2
BTK	CTIORF30 (EMSY)	CALR	CARD11	CASP8	CBFB	CBL	CCND1	CCND2
CCND3	CCNE1	CD22	CD274 (PD-L1)	CD70	CD79A	CD79B	CD73	CDH1
CDK12	CDK4	CDK6	CDK8	CDKN1A	CDKN1B	CDKN2A	CDKN2B	CDKN2C
CEBPA	CHEK1	CHEK2	CIC	CREBBP	CRKL	CSF1R	CSF3R	CTCF
CTNNA1	CTNNB1	CUL3	CUL4A	CXCR4	CYP17A1	DAXX	DDR1	DDR2
DIS3	DNMT3A	DOT1L	EED	EGFR	EP300	EPHA3	EPHB1	EPHB4
ERBB2	ERBB3	ERBB4	ERCC4	ERG	ERRF1	ESR1	EZH2	FAM46C
FANCA	FANCC	FANCG	FANCL	FAS	FBXW7	FGF10	FGF12	FGF14
FGF19	FGF23	FGF3	FGF4	FGF6	FGFR1	FGFR2	FGFR3	FGFR4
FH	FLCN	FLT1	FLT3	FOXL2	FURIN	GABRA6	GATA3	GATA4
GATA6	GID4 (CFZORF39)	GNAS1	GNAI3	GNAQ1	GNAS2	GRM7	GRIK3B	H3F3A
HDAC9	HGF	HNF1A	HNF1B	HNF1C	ID3	IL11	IL12	IL6
IKBKE	IKZF1	INF4B	INF2	INF4	IRS1	JAK1	JAK2	JAZF
KLHL30	KDL5A	KIF5C	KIF14	KDR	KEP1	KEP2	KIF11	KLHL6
KMT2A (MLL)	KMT2D (MLL2)	KRAS	LTK	LYN	MAF	MAP2K1 (MEK1)	MAP2K2 (MEK2)	MAP2K4
MAP3K1	MAP3K13	MAPK1	MCL1	MDM2	MDM4	MED12	MEF2B	MEN1
MERTK	MET	MITF	MKNK1	MLH1	MPL	MRE11A	MSH2	MSH3
MSH6	MST1R	MTOR	MTOR	MYC	MYCL (MYCL1)	MYCN	MYD88	
NBN	NF1	NFE2L1	NFE2L2	NKX2-1	NKX2-2	NOTCH2	NOTCH3	
NPM1	NRAS	NTRK1	NTRK2	NTRK3	NTRK3	PALB2	PARK2	
PARP1	PARP2	PAX1	PAX5	PBRM1	PDM1 (PDI)	PDGFRA	PDGFRA	PDGFRB
PDK1	PIK3C2B	PIK3C2G	PIK3CA	PIK3CB	PIK3R1	PIM1	PMS2	POLD1
POLE	PPARG	PPP2R1A	PPP2R2A	PRDM1	PRKARIA	PRKCJ	PTCH1	PTEN
PTPN11	PTPRO	QKI	RAC1	RAD21	RAD51	RAD51B	RAD51C	RAD51D
RAD52	RAD54L	RAF1	RARA	RB1	RBM10	REL	RET	RICTOR
RNF43	ROS1	RPTOR	SDHA	SDHB	SDHC	SDHD	SETD2	SF3B1
SGK1	SMAD2	SMAD4	SMARCA4	SMARCB1	SMO	SNCAIP	SOCS1	SOX2
SOX9	SPEN	SPOP	SRC	STAG2	STAT3	STK11	SUFU	SYK
TBX3	TEK	TET2	TGFBR2	TIPARP	TNFAIP3	TNFRSF14	TP53	TSC1
TSC2	TYRO3	U2AF1	VEGFA	VHL	WHSC1 (MMSET)	WHSC1L1	WT1	XPO1
XRCC2	ZNF217	ZNF703						

Foundation Medicine 324 Genes

Select Rearrangements^{2,3}

Genes with select intronic regions for the detection of gene rearrangements, one gene with a promoter region and one non-coding RNA gene.

ALK	BCL2	BCR	BRAF	BRCA1	BRCA2	CD74	EGFR	ETV4
ETV5	ETV6	EWSR1	EZR	FGFR1	FGFR2	FGFR3	KIT	KMT2A (MLL)
MSH2	MYB	MYC	NOTCH2	NTRK1	NTRK2	NUTM1	PDGFRA	RAF1
RARA	RET	ROS1	RSPO2	SDC4	SLC34A2	TERC*	TERT (PROMOTER ONLY)**	
TPRSS2								

*TERC is non-coding RNA gene.
**TERT is gene with promoter region.

Next-Generation Sequencing Expanded NGS Gene List

Mutations (DNA)									
ABL1	BRD4	CRLF2	FOXO4	HOXC11	KLFA	MUC1	PAK3	RHOH	TAL2
ABL1	BTG1	DDI2	FOXL3	HOXC13	KLK2	MUTYH	PRTZ1	RNF213	TBL1XR1
ACKR3	BTK	DDIT3	GATA1	HOXD11	LASP1	MYCL (MYCL1)	PAK8	RRL10	TCEA1
AKT1	C11orf65	DNM2	GATA2	HOXD13	LMO1	NBN	POE40P	SEPT5	TCL1A
AMER1 (FAM1238)	CBLC	DNMT3A	GNA11	HRA5	LMO2	NDRG1	PHF6	SEPT6	TERT
AR	CD79B	E1F4A2	GPC3	IKBKE	MAFB	NKX2-1	PHOX2B	SFPQ	TFE3
ARAF	CDH1	ELF4	HEY1	INHBA	MAX	NONO	PIK3CG	SLC45A3	TFPT
ATP2B3	CDK12	ELN	HIST1H3B	IRS2	MECOM	NOTCH1	PLG1	SMARCA4	THRAP3
ATRX	CDKN2B	ERCC1	HIST1H4I	JUN	MED12	NRAS	PMS1	SOC31	TLX3
BCL11B	CDKN2C	ETV4	HLF	KAT6A (MYST3)	MKL1	NUMA1	POU5F1	SOX2	TMPS2
BCL2	CEBPA	FAM66C	HMG2P46	KAT6B	MLL211	NUTM2B	PPP2R1A	SPOP	UBR5
BCL2L2	CHCHD7	FANCF	HNF1A	KCNJ5	MN1	OLIG2	PRF1	SRC	VHL
BCOR	CNOT3	FEV	HOXA11	KDM5C	MPL	OMD	PRKDC	SSX1	WAS
BCORL1	COL1A1	FOXL2	HOXA13	KDM6A	MSN	P2RY8	RAD21	STAG2	ZBTB16
BRD3	CDC6	FOXO3	HOXA9	KDSR	MTCP1	PFAH1B2	RECQL4	TAL1	ZRSR2
Mutations and Copy Number Variations (DNA)									
ABL2	BRCA2 ¹	COPB1	ESR1	FUS	KIT	MYB	PER1	RUNX1	TFG
ACSL3	BRIP1	CREB1	ETV1	GAS7	KLHL6	MYC	PICALM	RUNX1T1	TFRC
ACSL6	BUB1B	CREB3L1	ETV5	GATA3	MYO10	MYCN	PRK3A	SBD5	TGFB2
AF1	C11orf30 (EMS1)	CREB3L2	ETV6	GID4 (C17orf39)	KMT2D (MLL2)	MYD88	PRK31	SDC4	TLX1
AF3	C2orf44	CREBBP	EWSR1	GMP5	KMT2D (MLL2)	MYO10	PRK32	SDHAF2	TNFAIP3
AF4	CACNA1D	CRKL	EXT1	GNA13	KRAS	MYO10	PRK31	SDHB	TNFRSF1
AKAP9	CALR	CRTC1	EXT2	GNAQ2	KTNN1	MYO10	PRK31	SDHC	TNFRSF12
AKT2	CAMTA1	CRTC3	EZH2	GNA5	LGN	MYO10	PRK31	SDHD	TOP1
AKT3	CANM1	CSF1R	EZR	GOLGA5	LIG4	MYO10	PRK31	SEPT9	TRAF7
ALDH2	CARD11	CSF3R	FANCA	GOPC	LGR5	NCOA2	POT1	SET	TPM3
ALK	CAR5	CTCF	FANCC	GPHN	LHFPL	NCOA4	POLR2AF1	SETBP1	TPM4
APC	CAS5	CTLA4	FANCD2	GPR124	LIFR	NF1	PRARG	SETD2	TPR
ARFRP1	CASP8	CTNNA1	FANCE	GRIN2A	LPP	NF2	PRCC	SEB1	TRAF7
ARHGAP26	CBAF2T3	CTNNB1	FANGC	GSK3B	LRR13	NF2	PRDM1	SH2B3	TRIM26
ARHGGEF12	CBFB	CYLD	FANCL	H3F3A	LRR18	NFB	PRDM1	SH3GL1	TRIM27
ARID1A	CBL	CYP2D6	FAS	H3F3B	LYL1	NFKB2	PRKAR1A	SLC3	TRIM27
ARID2	CBFB	FBXO11	HERPUD1	MAF	NFKB1A	PRKAR1A	PRKX	SMAD4	TRIP11
ARNT	CDC6	DDR2	FBXW7	HGF	MALT1	NIN	PSIP1	MAD2	RAP
ASPSR1	CNNB1P1	DDX10	FCLRL4	HIP1	MAML2	NOTCH2	PTCH1	MARCB1	SC1
ASXL1	CCND1	DDX5	FGF10	HMG1	MAP2K1	NPM1	PTEN	SMARCA4	TSC2
ATF1	CCND2	DDX6	FGF14	HMG2	MAP2K2	NR4A3	PTPN11	SMD4	TSNR
ATIC	CCND3	DEK	FGF19	HNRNP2B1	MAP2K4	NSD1	PTPRC	SNX29	TTL
ATM	CCNE1	DICER1	FGF23	HOOK3	MAP3K1	NTS2	RABEP1	SOX10	UZAF1
ATP1A1	CD274 (PDL1)	DDIT1	FGF3	HSP90AA1	MLL1	NTRK1	RAC1	SPECC1	USP6
ATR	CD74	FGF4	FGF4	HSP90AB1	MDM2	NTRK2	RAD50	SREN	VEGFA
AURKA	CD79A	ECTZL	FGF6	IDH1	MDM4	NTRK3	RAD51	SRCAP3	VEGFB
AURKB	CD73	EGFR	FGFR1	IDH2	MDM2	NUP214	RAD51B	SRSF2	VTI1A
AXIN1	CDH11	ELK4	FGFR1OP	IGF1R	MEF2B	NUP93	RAF1	SRSF3	WHSC1
AXL	CDK4	ELL	FGFR2	IKZF1	MEN1	NUP98	RALGDS	SS18	WHSC1L1
BAP1	CDK6	EML4	FGFR3	IL2	MET (cMET)	NUTM1	RANBP17	SS18L1	WIF1
BARD1	CDK8	EP300	GFGR4	IL21R	MITF	PALB2	RAP1GDS1	STAT3	WSP3
BCL10	CDKN1B	EPHA3	FH	IL6ST	MLF1	PAX3	RARA	STAT4	WRN
BCL11A	CDKN2A	EPHA5	FHIT	IL7R	MLH1	PAX5	RB1	STAT5B	WT1
BCL2L11	CDX2	EPH81	FIP1L1	IRF4	MLL1	PAX7	RBM15	STIL	WTR1
BCL3	CHEK1	EP515	FLCN	ITK	MLL210	PBRM1	REL	STK11	XPA
BCL6	CHEK2	ERBB2 (HER2)	FLI1	JAK1	MLL3	PBX1	RET	SUFU	XPC
BCL7A	CHIC2	ERBB3 (HER3)	FLT1	JAK2	MLL4	PCM1	RICTOR	SUZ12	XPO1
BCL9	CHN1	ERBB4 (HER4)	FLT3	JAK3	MLL26	PCSK7	RM2	SYK	YWHAE
BCR	CIC	ERIC1	FLT4	JAZF1	MXN1	PDCD1 (PD1)	RNF43	TAF15	ZMYM2
BR3	CITF	ERCC2	FNBP1	KDM5A	MIE11A	PDCD1 (PD1)	RNF43	TCF12	ZNF217
BLM1	CLP1	ERCC3	FOXK1	KDR (VEGFR2)	MSH2	PDCD9	RNF22	TCF3	ZNF331
BMP81A	CLTC	ERCC4	FOXO1	KEAP1	MSH6	PDCFR	RPL5	TCF7L2	ZNF384
BRAF	CLTCL1	ERCC5	FOXP1	NIAA1549	MSI2	PDCFRB	RPN1	TEF1	ZNF521
BRCA1 ¹	CNPB	ERG	FUBP1	KIF5B	MTOR	PDK1	RPTOR	TET2	ZNF703
	CNTRL							TFEB	
Gene Fusions (RNA)									Variant Transcripts (RNA)
ALK	BRAF	NTRK1	NTRK2	NTRK3	RET	ROS1	RSPO3	EGFR vIII	MET Exon 14 Skipping

¹ May not be available for Medicare patients. Medicare reimburses BRCA1-2 for breast and ovarian cases only. Next-Generation Sequencing may not be available in New York State. For testing available in New York, please view the online New York Profile Menu (www.CarismolecularIntelligence.com/solid_tumors-NY).

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 Int: 00 41 21 533 53 00 | EUCustomerServices@carisls.com



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Tumor Profiling Services from Caris Molecular Intelligence®

	MI PROFILE™ Multi-platform, solid tumor biomarker analysis for therapeutic decision support	Next-Generation Sequencing Next-Generation Sequencing for additional biomarker analysis
Chemotherapy	✓	-
Immunotherapy	✓	-
Targeted Therapy	✓	✓
Protein Expression via IHC	✓	-
DNA Analysis via Pyro Sequencing	✓	-
DNA/RNA Analysis via Fragment Analysis	✓	-
Molecular Analysis via Next-Generation Sequencing	592 Genes	592 Genes
Clinical Support/Intit	✓	✓

Caris Life Sciences

592 Genes

Technical Specifications

Sufficient tumor must be present to complete all analysis. If you have any questions, please contact Client Services at (888) 979-8669.

Technical Information	IHC	CISH	FISH
Sample Requirements <i>(see request for full details)</i>	1 unstained slide at 4µm thickness from FFPE block, with evaluable tumor present, per IHC test	1 unstained slide at 4µm thickness from FFPE block, with at least 20-100 evaluable tumor cells present, per CISH test	2 unstained slides at 4µm thickness from FFPE block, with at least 100 evaluable cells present and 10% tumor, per FISH test
Sensitivity/Specificity	>95%	>95%	>95%

Technical Information	Next-Generation Sequencing	
	Mutations and Copy Number Variations (DNA)	Fusions (RNA)
Sample Requirements	FFPE block or 15 unstained slides with a minimum of 20% malignant origin. Needle biopsy is also acceptable (4-6 cores).	FFPE block or 2-5 unstained slides with a minimum of 20% malignant origin. Needle biopsy is also acceptable (4-6 cores).
Tumor Enrichment	Microdissection performed on all cases resulting in ~25% increase in tumor nuclei and enhances detection of minor clonal variants	
Amount of DNA Required	200ng input (50ng)	
PPV	>99%	>98%
Sensitivity	> 99% for base substitutions at ≥ 5% mutant allele frequency; > 99% for indels at ≥ 5% mutant allele frequency; >95% for copy number variations (amplifications ≥ 8 copies)	>91%
Average Depth of Coverage (DNA) Average Depth/Count (RNA)	>750X	>30,000 Unique RNA Fragments
Number of Genes	592 genes	10 genes

Gene List

Guardant360 CDx is indicated to provide tumor mutation profiling for advanced cancer patients with any solid malignant neoplasm. Guardant360 CDx report contains both professional services, which includes 74 genes, in addition to the FDA-approved report, which includes 55 genes.

Point Mutations (SNVs) and Deletion Variants (Indels) (74 Genes)						Amplifications (18 Genes)		Fusions (6 Genes)
AKT1	CDH1	FGFR2	KRAS	NPM1	RIT1	AR	FGFR1	ALK
ALK	CDK4	FGFR3	MAP2K1	NRAS	ROS1	BRAF	FGFR2	FGFR2
APC	CTNNB1	GATA3	MAP2K2	TRK1	SMAD4	CCND1	KIT	FGFR3
AR	DKT2	GNAS	MAPK1	TRK3	SMO	CCND2	KRAS	NTRK1
ARAF	CDKN2A	GNAQ	MAPK3	PDGFRA	STK11	CCNE1	MET	RET
ARID1A	CTNNB1	GNAS	MET	PIK3CA	TERT [^]	CDK4	MYC	ROS1
ATM	DDR2	HNF1A	MTH1	PTEN	PSMA	CDK5	PDGFRA	
BRAF	EGFR	HRAS	MPL	PTPN11	TSC1	EGFR	PIK3CA	
BRCA1	ERBB2	IDH1	MTOR	RAF1	VHL	ERBB2	RAF1	
BRCA2	ESR1	IDH2	MYC	RB1				
CCND1	EZH2	JAK2	NF1	RET				
CCND2	FBXW7	JAK3	NFE2L2	RHEB				
CCNE1	FGFR1	KIT	NOTCH1	RHOA				

Guardant Health
74 Genes

Critical or all exons* completely sequenced and all four major classes of alterations

NSCLC guideline-recommended genes shown in bold / *Exons selected to maximize detection of known somatic mutations / ^ Includes TERT promoter region

Traditional Abstraction of Gene Mutations?

- Site Specific Data Items (SSDIs) take years to approve
 - Long after testing and clinical use have become standards of clinical care
- Registrars do not have time to review and manually code hundreds of gene mutations per case
- Obtaining test results directly from sequencing providers will be much more efficient and complete



Central Registry Infrastructure Needed to Capture Genomic Test Data

Moving Beyond the Limitations

Commercial Laboratory NGS Panel Testing and Reporting

Clinical Report

- Specific gene mutations from tumor tissue
- Suggestions for FDA approved targeted agents and clinical trials
- May or may not report variants of unknown significance

Raw Data used to Generate Clinical Report

- Sequencer -> FastQ -> BAM -> VCF -> Clinical Report
- Clinical report based upon current knowledge of mutation variants
- FastQ and BAM files contain information that may prove important in future
- At minimum, BAM files important for surveillance



FOUNDATION ONE

Patient Name: Patient, Test
 Report Date: 09 December 2016
 Tumor Type: Breast carcinoma (NOS)

Date of Birth	12 December 1942	Medical Facility	ABC Oncology		
Sex	Female	Ordering Physician	Smith, John	Specimen Received	01 December 2016
FMI Case #	SMP60684	Additional Recipient	Not Given		
Medical Record #	12345678	Medical Facility ID #	200313		
Specimen ID	Not Given	Pathologist	Public, John Q.	Specimen Type	Slide

ABOUT THE TEST:

FoundationOne™ is a next-generation sequencing (NGS) based assay that identifies genomic alterations within hundreds of cancer-related genes.

PATIENT RESULTS

4 genomic findings

2 therapies associated with potential clinical benefit

0 therapies associated with lack of response

10 clinical trials

TUMOR TYPE: BREAST CARCINOMA (NOS)

Genomic Alterations Identified[†]

PIK3CA E545K
ATM T2333fs*40
BCL2L1 amplification – equivocal*
MYST3 amplification – equivocal*

Additional Disease-relevant Genes with No Reportable Alterations Identified[†]

ERBB2

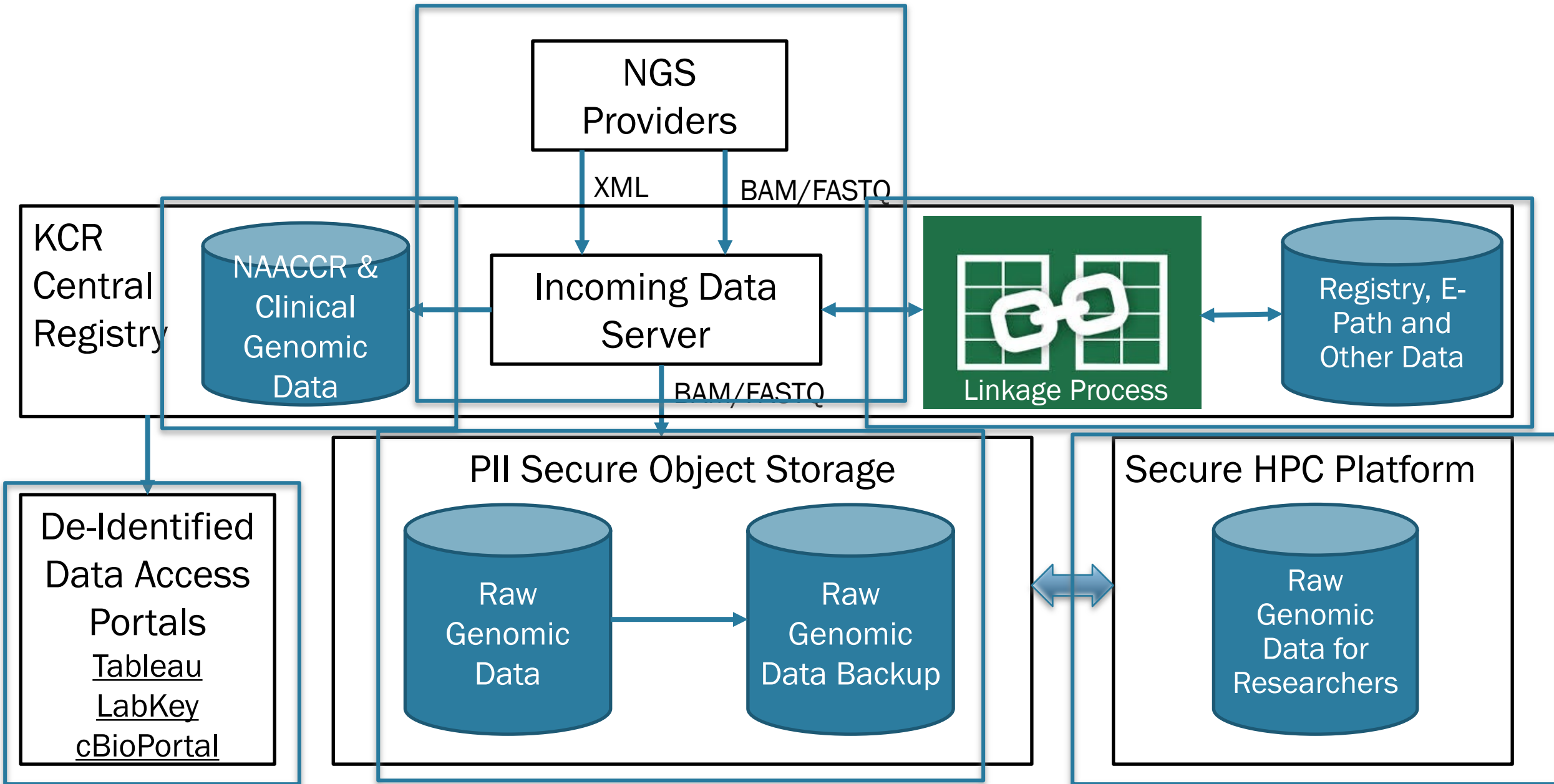
[†] For a complete list of the genes assayed and performance specifications, please refer to the Appendix
^{*} See Appendix for details

THERAPEUTIC IMPLICATIONS

Genomic Findings Detected	FDA-Approved Therapies (in patient's tumor type)	FDA-Approved Therapies (in another tumor type)	Potential Clinical Trials
<i>PIK3CA</i> E545K	Everolimus	Temsirolimus	Yes, see clinical trials section
<i>ATM</i> T2333fs*40	None	None	Yes, see clinical trials section
<i>BCL2L1</i> amplification - equivocal	None	None	None
<i>MYST3</i> amplification - equivocal	None	None	None

Note: Genomic alterations detected may be associated with activity of certain FDA-approved drugs; however, the agents listed in this report may have little or no evidence in the patient's tumor type. Neither the therapeutic agents nor the trials identified are ranked in order of potential or predicted efficacy for this patient, nor are they ranked in order of level of evidence for this patient's tumor type.

Genomic Data Flow into the Central Registry



KCR/MCC cBioPortal for Cancer Genomics

- I. The cBioPortal for Cancer Genomics is an open-access, open-source resource for interactive exploration of multidimensional cancer genomics data sets. The goal of cBioPortal is to significantly lower the barriers between complex genomic data and cancer researchers by providing rapid, intuitive, and high-quality access to molecular profiles and clinical attributes from large-scale cancer genomics projects, and therefore to empower researchers to translate these rich data sets into biologic insights and clinical applications.
- II. Provide representative, de-identified, population-based data from Kentucky cancer patients annotated with high quality KCR data





Combined Study

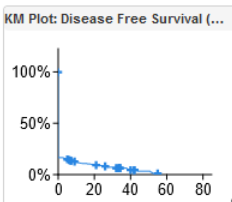
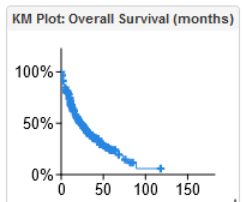
This combined study contains samples from 14 studies

Click gene symbols below or enter here Query

Summary Clinical Data

Selected: 298 patients | 304 samples

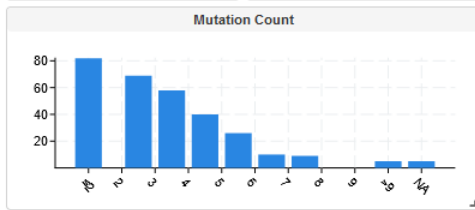
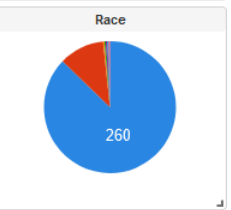
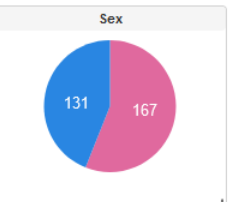
Custom Selection Charts Groups



Gene	# Mut	#	Freq
TP53	267	233	77.2%
KRAS	99	98	32.5%
STK11	46	46	15.2%
CDKN2A	33	32	10.6%
SMARCA4	31	31	10.3%
KEAP1	27	27	8.9%
ARID1A	26	25	8.3%
NF1	27	24	7.9%
RBM10	22	22	7.3%
PIK3CA	21	20	6.6%
LRP1B	22	20	6.6%

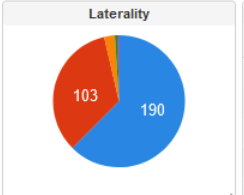
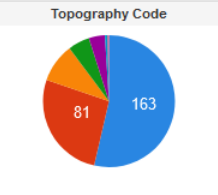
Gene	# SV	#	Freq
ALK	4	4	1.4%
STK11	4	4	1.4%
BRCA1	3	3	1.0%
NF1	3	3	1.0%
LRP1B	3	3	1.0%
CDC73	3	3	1.0%
RB1	2	2	0.7%
RET	2	2	0.7%
ROS1	2	2	0.7%
APC	1	1	0.3%
CD74	1	1	0.3%

Gene	CytoBand	CNA	#	Freq
MYC	8q24.21	AMP	34	11.5%
SOX2	3q26.33	AMP	28	9.5%
RICTOR	5p13.1	AMP	27	9.2%
NKX2-1	14q13.3	AMP	24	8.1%
PIK3CA	3q26.32	AMP	21	7.1%
FGF10	5p12	AMP	19	6.4%
NFKBIA	14q13.2	AMP	19	6.4%
TERC	3q26.2	AMP	19	6.4%
PRKCI	3q26.2	AMP	17	5.8%
CCND1	11q13.3	AMP	16	5.4%
FGF19	11q13.3	AMP	16	5.4%

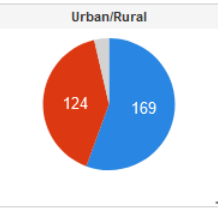
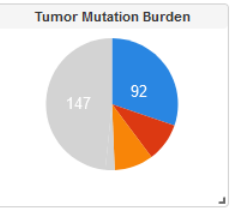
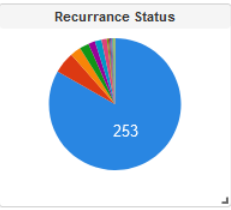
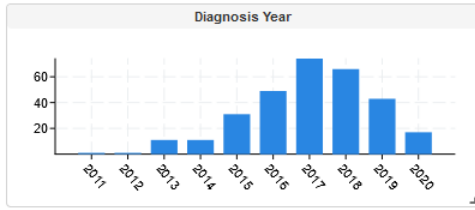
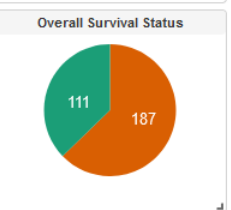
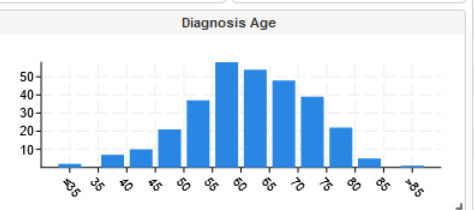
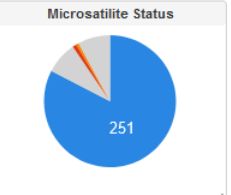


Histology	#	Freq
ADENOCARCINOMA, NOS	161	53.0%
SQUAMOUS CELL CARCINOMA	67	22.0%
NON SM CELL CARCINOMA	19	6.3%
LARGE CELL NEUROENDOCRINE	9	3.0%
MUCINOUS ADENOCARCINOMA	7	2.3%
SMALL CELL CARCINOMA, NOS	7	2.3%
ACINAR CELL CARCINOMA	4	1.3%
ACINAR CELL CYSTADENOCARCINOMA	4	1.3%
NEUROENDOCRINE CARCINOMA	3	1.0%
PAPILLARY ADENOCARCINOMA	3	1.0%
PLEOMORPHIC CARCINOMA	3	1.0%

Histology Code	#	Freq
8140	161	53.0%
8070	67	22.0%
8046	19	6.3%
8013	9	3.0%
8041	7	2.3%
8480	7	2.3%
8550	4	1.3%
8551	4	1.3%
8022	3	1.0%
8246	3	1.0%
8260	3	1.0%



Best Stage Group	#	Freq
Stage IV	87	28.6%
Clinical Stage IVB	34	11.2%
Pathologic Stage IVB	24	7.9%
Stage IB	23	7.6%
Stage IIIB	21	6.9%
Pathologic Stage IVA	18	5.9%
Stage IA	18	5.9%
Stage IIIA	18	5.9%
Clinical Stage IVA	8	2.6%
Stage IIB	8	2.6%
Clinical Stage IIIA	6	2.0%



Transmission and Storage Requirements

- Secure FTP between central registry and sequencing provider
 - Push or pull
- Molecular data files must contain linkage identifiers
 - Patient: Last Name, First Name, Date of Birth, SSN, Medical Record Number
 - Case: Diagnosis (Site/Histology), Diagnosis Date, Path Report Number (Specimen), Specimen Date
- Data storage needed

Data Type	Average Size	Storage for 1000 Records
XML (Mutations)	< 1 Mb	< 1 GB
PDF (Clinical Reports)	< 1 Mb	< 1 GB
BAM (Processed Raw Data)	2.3 GB (+10MB Index)	~3 TB
FASTQ (WES Raw Data)	20-40GB	20-40 TB

Other Resource Considerations

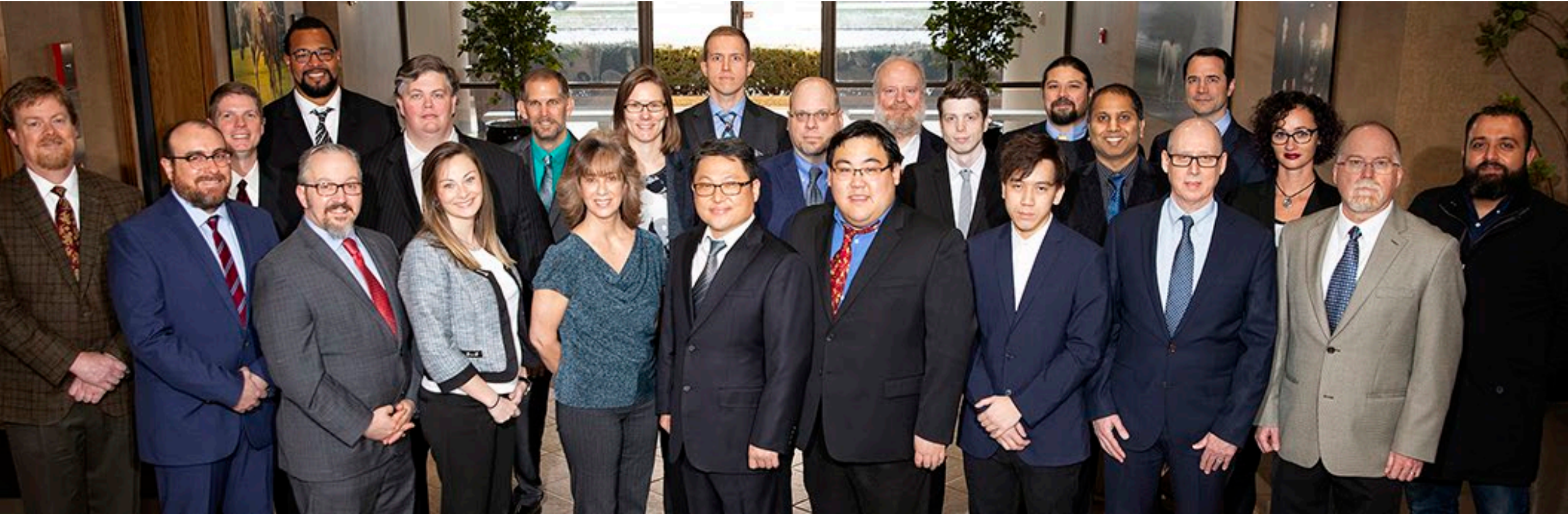
■ Legal

- Data Use Agreements with Sequencing Providers
- Hospital Agreements (permission to send results to registry)

■ Staffing

- Technical infrastructure development
- PII security (protecting germline sequencing)
- Bioinformatics support
- Cancer registrars for reviewing and linking genomic reports with path reports and registry data

Acknowledgements: KCR/Markey Informatics Teams



Questions/Discussion

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